

ANTITUMOR AGENTS, 93.¹ BRUCEANOL C, A NEW CYTOTOXIC QUASSINOID FROM *BRUCEA ANTIDYSENTERICA*

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Kupchan and associates (1) isolated eight quassinoids, including the anti-leukemic bruceantin that reached phase II clinical trials at the National Cancer Institute, from the Ethiopian *Brucea antidysenterica* Mill. (Simaroubaceae).

Recently we reported on the isolation and structural elucidation of four new antileukemic quassinoids, bruceantinoside A and B (2), bruceanol A and B (3), and cytotoxic antileukemic alkaloids (4,5) from the stem of *B. antidysenterica*. We now describe the isolation and characterization of a new, potent, cytotoxic quassinoid, bruceanol C [1], from this same plant.

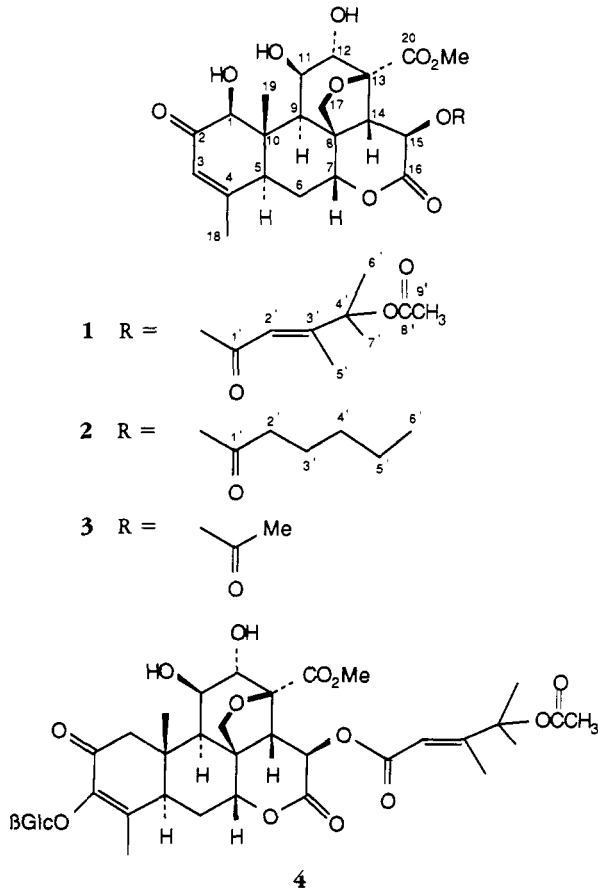
Bruceanol C [1] was obtained as a colorless amorphous powder. The ¹H-nmr (Table 1) signals for H-1, H-3, H-7, H-12, H-15, Me-4, Me-10, and OMe of 1 were coincident with those of bruceanol B [2] and isobruceine B [3]. On the other hand, the signals for H-2', Me-3', Me-4', and OAc-4' of 1 were very similar to those of yadanzioside K [4] (6). Two singlets for Me-4' at 1.51 and 1.53 ppm are due to the OAc substitution at 4' position of the C-15 side chain. The ¹³C-nmr signals for C-1~Me-10 of 1 coincided with those of 2 and 3. On the other hand, the signals for C-1'~C-9' of 1 were coincident with those of 4. These

data led to the assignment of structure 1 for bruceanol C. The ir spectrum also supported this structure. From this structure, the molecular formula should be C₃₀H₃₈O₁₃ (mol wt 606); however, eims showed the highest peak at *m/z* 546 instead of *m/z* 606. High resolution eims showed a peak at *m/z* 546.2105, which corresponds to C₂₈H₃₄O₁₁ ([M - C₂H₄O₂]⁺), indicating the elimination of HOAc from 1.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—Melting points were determined on an MRK air-bath-type melting point apparatus and were uncorrected. Specific rotations were obtained on a YANAKO OR-50D polarimeter (L = 0.1 dm). Ir and uv spectra were recorded on a JASCO IR-810 spectrometer and a Hitachi 320-S spectrometer, respectively. ¹H-nmr and ¹³C-nmr spectra were determined on a JEOL JNM-GX400 (400 MHz for ¹H nmr and 100.40 MHz for ¹³C nmr) in CDCl₃ using TMS as an internal standard, and the results are shown in Tables 1 and 2, respectively. The assignments of the carbon signals were made by off-resonance decoupling. Mass spectra were recorded on a Hitachi M80 instrument. Si gel (Merck, type 60, 70-230 mesh) was used for cc, and precoated Si gel plates (Merck, 60F-254, 0.25 mm) were used for analytical tlc. Detection of components was made by use of an uv lamp. Analytical hplc was performed on a Waters Associates Model ALC/GPC 244 liquid chromatograph equipped with Radial Pak Liquid Chromatography Cartridge (8NVC18). Preparative hplc was done on a Gilson preparative liquid chromatograph equipped with an M&S PACK C18-A column (20 × 250 mm).

¹For part 92, see Y.C. Wu, S.T. Lu, J.J. Chang, and K.H. Lee, *Phytochemistry*, in press.

TABLE 1. $^1\text{H-nmr}$ Spectra of Quassinoids **1**, **2**, **3**, and **4**.

Proton	Compound			
	Bruceanol C [1] ^a	Bruceanol B [2] ^a	Isobruceine B [3] ^b	Yadanzioside K [4] ^c
H-1	4.29 s	4.28 br s	4.26 s	f
H-3	6.10 br s	6.1 br s	6.11 br s	f
H-7	4.8 m	4.75 br s	4.75 m	f
H-11	4.75 br s	4.54 br s	4.75 m	f
H-12	4.15 s	4.18 br s	4.12 br s	f
H-15	6.3 d (13) ^d	6.29 d (13) ^d	6.30 d (13) ^d	6.85 d (13) ^d
Me-4	1.96 s	1.96 s	1.95 s	2.04 s
Me-10	1.18 s	1.19 s	1.20 s	1.68 s
OMe	3.79 s	3.83 s	3.83 s	3.89 s
H-2'	5.78 s	e	2.08 s	6.06 s
Me-3'	2.13 s	e	—	2.26 s
Me-4'	1.51 s	e	—	1.42 s
	1.53 s	e	—	1.47 s
OAc-4'	2.02 s	e	—	1.98 s

^aMeasured at 400 MHz in CDCl_3 .^bMeasured at 400 MHz in CDCl_3 by Moretti *et al.* (7).^cMeasured at 90 MHz in $\text{C}_5\text{D}_5\text{N}$ by Sakaki *et al.* (6).^dCoupling constant in Hz.^eThe signals of H-2' ~ H-6' were found at 1.3 ppm (m) and 0.91 ppm (t, $J = 7$ Hz).^fNot measured.

TABLE 2. ¹³C-nmr Spectra of Quassinoids 1, 2, 3, and 4.

Carbon	Compound			
	Bruceanol C [1] ^a	Bruceanol B [2] ^a	Isobruceine B [3] ^b	Yadanzioside K [4] ^c
C-1	80.5 d	80.6 d	81.3 d	51.1 t
C-2	196.9 s	196.9 s	197.6 s	193.6 s
C-3	124.2 d	124.3 d	124.5 d	148.0 s
C-4	163.1 s	162.9 s	162.6 s	146.6 s
C-5	43.5 d	43.5 d	43.4 d	43.4 d
C-6	28.5 t	28.5 t	28.2 t	29.3 t
C-7	81.2 d	81.1 d	81.7 d	83.4 d
C-8	45.5 s	45.7 s	45.8 s	46.0 s
C-9	42.6 d	42.7 d	42.4 d	41.9 d
C-10	47.5 s	47.5 s	47.7 s	40.8 s
C-11	72.4 d	72.5 d	74.3 d	73.0 d
C-12	75.7 d	75.8 d	75.1 d	75.9 d
C-13	82.2 s	82.8 s	81.7 s	82.6 s
C-14	51.3 d	51.5 d	52.3 d	50.2 d
C-15	66.4 d	66.5 d	67.8 d	68.4 d
C-16	167.0 s	166.9 s	167.6 s	168.0 s
C-17	73.3 t	73.3 t	73.0 t	73.5 t
C-18	22.6 q	22.5 q	22.4 q	15.2 q
C-19	11.6 q	11.5 q	11.3 q	15.8 q
C-20	172.1 s	172.4 s	169.5 s	171.2 s
OMe	53.1 q	52.8 q	49.8 s	52.5 q
C-1'	165.2 s	171.9 s	170.7 s	165.7 s
C-2'	111.9 d	33.6 t	20.5 q	113.5 d
C-3'	169.5 s	31.2 t	—	169.5 s
C-4'	82.7 s	24.2 t	—	82.3 s
C-5'	14.6 q	22.2 t	—	14.3 q
C-6'	26.0 q	13.8 q	—	25.8 q
C-7'	26.3 q	—	—	26.3 q
C-8'	164.7 s	—	—	163.4 s
C-9'	21.6 q	—	—	21.4 q

^aMeasured at 100.40 MHz in CDCl₃.

^bMeasured at 22.63 MHz in CDCl₃ by Moretti *et al.* (7).

^cMeasured at 22.5 MHz in C₅D₅N by Sakaki *et al.* (6).

CHROMATOGRAPHY OF THE CHCl₃ FRACTIONS.—The crude CHCl₃ fraction (266 g), which was part of the CHCl₃ extract of the ground wood of *B. antidiysenterica* (1918 kg) reported previously (1), was subjected to cc on Si gel (3 kg, 10×90 cm) and eluted first with CHCl₃ and then with increasing amounts of MeOH in CHCl₃ to yield 19 fractions. Concentration of fraction 11 by evaporation of the solvent afforded a brown gum (7.55 g).

ISOLATION OF BRUCEANOL C [1].—The foregoing brown gum (7.55 g) was subjected to analytical hplc using a mixed solvent of MeOH-H₂O (1:1, v/v), and a new peak was found by comparing the retention times with those of known quassinoids and alkaloids. Therefore, the gum was subjected twice to preparative hplc

using an M&S PACK C18-A column (20 mm×25 cm), a mixed solvent of MeOH-H₂O (3:1, v/v; flow rate 2 ml/min), and Gilson Model 111B uv detector at 254 nm. This led to a yellow, resinous substance (397 mg) that was purified by repeated recrystallization from EtOAc/CH₂Cl₂/Et₂O to afford the pure compound 1 (34 mg, 0.0000017%). This compound was characterized as follows:

BRUCEANOL C [1].—Colorless, amorphous powder, mp 125–127°; [α]_D²⁵+38° (c=0.24, EtOH); uv λ max (EtOH) 225 nm (ε 14000); ir (KBr) 3560 and 3450 (OH), 1735 (ester and lactone C=O), 1675 (α,β-unsaturated C=O), 1645 (C=C), 1385 and 1375 (*gem*-demethyl), 1262 (acetate), 1060 (OH), 965 (C=C) cm⁻¹; ¹H nmr and ¹³C nmr see Tables 1 and 2, respec-

tively; hreims m/z 546.2105 (calcd for $C_{28}H_{34}O_{11}$ $[M - C_2H_4O_2]^+$, 546.2100).

BIOLOGICAL ACTIVITY.—The in vitro tissue culture cytotoxicity assay was carried out according to standard National Cancer Institute procedure (8).² Bruceanol C demonstrated potent cytotoxicity against human KB, A-549 lung carcinoma, and HCT-8 colon tumor as well as murine P-388 lymphocytic leukemia with ED₅₀ values of <0.04, 0.48, <0.40, and 0.56 μ g/ml, respectively.

ACKNOWLEDGMENTS

This investigation was supported by a grant from the National Cancer Institute CA 17625 (K.H. Lee). The authors thank Drs. John Douros and Matthew Suffness, Division of Cancer Treatment, National Cancer Institute, for their interest and assistance in providing the plant extract; Drs. Yung-Chi Cheng and Jer-Jang Chang and Mr. Mike Fisher, School of Medicine, University of North Carolina at Chapel Hill, for bioassay; Miss Shigeo Miki and Mr. Akiharu Zushi, Meijiseika Kaisha Ltd., for hreims and ¹H and ¹³C nmr, respectively; and Miss T. Sai, Kobe Women's College of Pharmacy, for fdms.

LITERATURE CITED

1. S.M. Kupchan, R.W. Britton, J.A. Lacadie, M.F. Ziegler, and C.W. Sigel, *J. Org. Chem.*, **40**, 648 (1975).
2. M. Okano, K.H. Lee, I.H. Hall, and F.E. Boettner, *J. Nat. Prod.*, **44**, 470 (1981).
3. M. Okano, N. Fukamiya, T. Aratani, M. Ju-ichi, and K.H. Lee, *J. Nat. Prod.*, **48**, 972 (1985).
4. N. Fukamiya, M. Okano, T. Aratani, K. Negoro, A.T. McPhail, M. Ju-ichi, and K.H. Lee, *J. Nat. Prod.*, **49**, 428 (1986).
5. N. Fukamiya, M. Okano, T. Aratani, K. Negoro, Y.M. Lin, and K.H. Lee, *Planta Med.*, **2**, 140 (1987).
6. T. Sakaki, S. Yoshimura, T. Tsuyuki, T. Takahashi, T. Honda, and T. Nakanishi, *Bull. Chem. Soc. Jpn.*, **59**, 3541 (1986).
7. C. Moretti, J. Polonsky, M. Vuilhorgne, and T. Prange, *Tetrahedron Lett.*, **23**, 647 (1982).
8. R.I. Geran, N.H. Greenberg, M.M. MacDonald, A.M. Schumacher, and B.J. Abbott, *Cancer Chemother. Rep., Part 3*, **3**, 1 (1972).

Received 8 September 1987

²K.H. Lee, Y.M. Lin, T.S. Wu, D.C. Zhang, T. Yamagishi, T. Hayashi, I.H. Hall, J.J. Chang, R.Y. Wu, and T.H. Yang, *Planta Med.* (in press).